



**LYMPHOGRAM®**

Ref: CYT-C001



*For In Vitro Diagnostic use*

### **INTENDED USE**

LYMPHOGRAM® is a three-color direct immunofluorescence reagent for use in flow cytometry designed to simultaneously determine in peripheral blood, bone marrow and other body fluids the major lymphocyte subpopulations, including the total number of T lymphocytes (CD3+), B lymphocytes (CD19+) and natural killer cells (CD3-CD56+) as well as helper/inducer (CD3+CD4+) and suppressor/cytotoxic (CD3+CD8+) T lymphocyte subsets.

### **SUMMARY AND EXPLANATION**

Flow Cytometry is a powerful tool for the analytical and quantitative characterization of cells which provides rapid, quantitative and multiparametric analysis of heterogeneous cell populations on a cell-by-cell basis. Flow cytometry is performed on cells in liquid suspension that have been incubated with fluorescently-labeled antibodies directed against specific cellular proteins. The relative fluorescence intensity of the positive cells indicates the amount of antibody bound to specific binding sites on the cells, and therefore provides a relative measure of antigen expression.

Human lymphocytes may be classified in three main populations according to their biological function and their cell surface antigen expression: T lymphocytes, B lymphocytes and natural killer (NK) cells. T lymphocytes (CD3+), the precursors of which originate in the bone marrow and then migrate and mature in the thymus, can be subdivided as well in functionally different populations. The most clearly defined of these are helper/inducer T cells (CD3+CD4+) and suppressor/cytotoxic T cells (CD3+CD8+). T cells produce no antibodies and are the mediators of cell immunity. B lymphocytes (CD19+) are the producers of antibodies, they mediate humoral immunity particularly effective against toxins, whole bacteria, and free viruses. NK cells (CD3-CD56+) mediate cytotoxicity against certain tumors and virus-infected cells. NK-mediated cytotoxicity does not require class I or class II major histocompatibility complex (MHC) molecules to be present on the target cell.

LYMPHOGRAM® recognizes the antigens CD3, CD19, CD56, CD4 and CD8 present in the different lymphocyte subsets, and can therefore be used in the characterization studies for immunophenotyping of lymphocytes. These studies are widely applied for monitoring of the immunologic status of post-transplant patients and in the characterization and follow-up of immunodeficiencies, autoimmune diseases, leukemia etc<sup>(1-3)</sup>.

### **PRINCIPLES OF THE PROCEDURE**

Flow cytometry (FC) is an innovative technology by means of which different cell characteristics are simultaneously analyzed on a single cell basis. This is achieved by means of hydrodynamic focusing of cells that pass aligned one by one in front of a set of light detectors; at the same time they are illuminated by a laser beam. The interaction of the cells with the laser beam generates signals of two different kinds: those generated by dispersed light (FSC/SSC), which mainly reflects the size of the cell and its internal complexity, and those related to the emission of light by the fluorochromes present in the cell. These signals become electric impulses which are amplified and registered as digital signals to be processed by a computer.

When the reagent is added to the sample, the mixture of fluorochrome-labelled antibodies present in the reagent bind specifically to the antigens they are directed against, allowing the detection by FC of the different lymphoid subsets.

The erythrocyte population, which could hinder the detection of the target population, is eliminated by the use of a red blood cell lysing solution previous to acquire the sample on the cytometer. The use of Quicklysis™ (CYT-QL-1) erythrocyte lysing solution is recommended, since it requires no further washing step and contains no fixative, therefore minimizing the handling of the sample and avoiding the cell loss associated to the centrifuge process.<sup>(4, 5)</sup>

The different lymphocyte subsets count is generally expressed as the number of positive cells per microliter of sample (absolute counts), or as the percentage of positive cells per lymphocytes or leucocytes present in the sample which can itself be determined by FC based on its typical pattern of FSC/SSC (size/granularity or complexity). Because each flow cytometer has different operating characteristics each laboratory must determine its optimal operating procedure.

### **REAGENT COMPOSITION**

LYMPHOGRAM® is provided in phosphate buffered saline with 0.1% sodium azide. It contains:

- Purified monoclonal CD8 antibody conjugated with fluorescein isothiocyanate (FITC), clone: UCH-T4, isotype: IgG2a
- Purified monoclonal CD19 antibody conjugated with fluorescein isothiocyanate (FITC), clone: HD37, isotype: IgG1
- Purified monoclonal CD3 antibody conjugated with R-phycoerythrin (PE), clone: 33-2-A3, isotype: IgG2a
- Purified monoclonal CD56 antibody conjugated with R-phycoerythrin (PE), clone: C5.9, isotype: IgG2b
- Purified monoclonal CD4 antibody conjugated with the R-phycoerythrin - cyanine 5 tandem (PECy5), clone: 13B8.2, isotype: IgG1

Purification: Affinity chromatography

Amount per 1,25 ml vial: 50 tests (25 µl LYMPHOGRAM® to 10<sup>6</sup> cells)

Reagents are not considered sterile.

### **STORAGE CONDITIONS**

The reagent is stable until the expiration date shown on the label, when stored at 2-8° C. The reagent should not be frozen or exposed to direct light during storage or during incubation with cells. Keep the reagent vial dry. Once opened, the vial must be stored in a vertical position to avoid any possible spillage.

### **WARNINGS AND RECOMMENDATIONS**

1. For in vitro diagnostic use.
2. This product is supplied ready to use. If it is altered by dilution or addition of other components, it will be invalidated for in vitro diagnostic use.
3. The reagent is stable until the expiration date shown on the label if it is properly stored. Do not use after the expiration date shown on the label. If the reagents are stored in conditions different from those recommended, such conditions must be validated by the user.

4. Alteration in the appearance of the reagent, such as the precipitation or discoloration indicates instability or deterioration. In such cases, the reagent should not be used.
5. It contains 0.1% sodium azide (CAS-Nr. 26628-22-8) as a preservative, but even so care should be taken to avoid microbial contamination of reagent or incorrect results may occur.
  - Sodium azide ( $\text{NaN}_3$ ) is harmful if swallowed (R22), if swallowed, seek medical advice immediately and show this container or label (S46).
  - Wear suitable protecting clothing (S36).
  - Contact with acids liberates very toxic gas (R32).
  - Azide compounds should be flushed with large volumes of water during disposal to avoid deposits in metal drains where explosive conditions may develop.
6. All patient specimens and materials with which they come into contact are considered biohazards and should be handled as if capable of transmitting infection<sup>(6)</sup>, and disposed according to the legal precautions established for this type of product. Also recommended is handling of the product with appropriate protective gloves and clothing, and its use by personnel sufficiently qualified for the procedures described. Avoid contact of samples with skin and mucous membranes. After contact with skin, wash immediately with plenty of water.
7. Use of the reagent with incubation times or temperatures different from those recommended may cause erroneous results. Any such changes must be validated by the user.

## PROCEDURE

### Material included

LYMPHOGRAM<sup>®</sup> is sufficient for 50 determinations (25  $\mu\text{L}$  reagent to  $10^6$  cells).

### Material required but not included

- 488 nm ion argon laser-equipped flow cytometer and appropriate computer hardware and software.
- Test tubes suitable for obtaining samples in the flow cytometer used. Usually tubes with a rounded bottom for 6 mL, 12x 75 mm are used.
- Automatic pipette (100 $\mu\text{L}$ ) and tips.
- Micropipette with tips.
- Chronometer
- Vortex Mixer
- Isotypic control reagent
- Quicklysis<sup>™</sup> lysing solution
- Wash buffer as phosphate buffered saline (PBS) pH 7.4 containing 0,1% sodium azide.
- Perfect-Count Microspheres<sup>™</sup> (CYT-PCM-50) necessary for determining absolute counts.

### Preparation

The protocol to process samples with LYMPHOGRAM<sup>®</sup> reagent depends on the procedure of the sample:

- Whole blood sample must be taken aseptically by means of a venipuncture<sup>(7, 8)</sup> in a sterilized tube for blood collection containing an appropriate anticoagulant (use of EDTA is recommended). The analysis requires one hundred (100)  $\mu\text{L}$  of the whole blood sample per tube, assuming a normal range of approximately 4 to  $10 \times 10^3$  leucocytes per  $\mu\text{L}$ . For samples with a high white blood cell count, dilute samples with PBS to obtain a concentration of cells approximately equal to  $1 \times 10^4$  cells/ $\mu\text{L}$ . Store the blood samples at 18-22°C until they are to be tested. It is advisable to test blood samples within the 24 hours after their extraction. Hemolyzed samples or samples with suspended cell aggregates should be rejected.
- Bone marrow sample should be pass 3 or 4 times through a syringe in order to disaggregate cell clumps. Perform a white blood cell count of the sample and dilute samples with PBS to obtain a concentration of cells approximately equal to  $1 \times 10^4$  cells/ $\mu\text{L}$ .
- Dilution is not required for other body fluid samples such as fine needle aspiration (FNA) or cerebrospinal fluid, which contain low lymphocyte counts.

1. Mix 100 $\mu\text{L}$  of blood or bone marrow sample with 25 $\mu\text{L}$  of LYMPHOGRAM<sup>®</sup>. In case of working with other body fluids with fewer cells, such as fine needle aspirations, cerebrospinal fluid, bronchoalveolar lavage, gastric lavage, etc, mix 200 $\mu\text{L}$  of sample with 25 $\mu\text{L}$  of LYMPHOGRAM<sup>®</sup>.

To evaluate the non-specific binding of the antibody, an appropriated isotype control tube can be prepared.

3. Incubate for 10 minutes at room temperature in the dark.
4. Add 2 ml of Quicklysis<sup>™\*</sup> erythrocyte lysing solution and incubate the sample for 10 minutes at room temperature in the dark.
5. Acquire directly on the flow cytometer within the first four hours after finishing the sample preparation. If the samples are not acquired immediately after preparation, they should be stored at 2-8°C in the dark. Calibration of the instrument must be done according to the manufacturer's advice. Before acquiring samples, adjust the threshold or discriminator to minimize debris and ensure populations of interest are included. Before acquiring the sample on the flow cytometer, mix the cells on the vortex at low speed to reduce aggregation.

\*Note: The use of other lysing solutions may require the elimination of the lysed red blood cells. Follow the manufacturer's recommended protocol of the lysing solution used.

If using Perfect-Count Microspheres<sup>™</sup> for calculating absolute counts, mix the Perfect-Count Microspheres<sup>™</sup> vial manually for 30-40 seconds (do not use Vortex) immediately prior to add to the sample. Using the reverse pipetting technique, add to each tube the same volume of Perfect-Count Microspheres<sup>™</sup> as the one used for the sample addition (100  $\mu\text{L}$  in case of blood or bone marrow sample and 200  $\mu\text{L}$  in case of other body fluids sample)

### Flow cytometry analysis

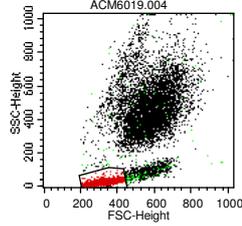
Check that the cytometer is correctly aligned and standardized for light dispersion (FSC/SSC on linear scale) and fluorescent intensity (FL1, FL2, FL3 FL4 on logarithmic scale) and that the right color compensation has been set following the instructions of the cytometer manufacturer.

Analysis of the LYMPHOGRAM<sup>®</sup> files becomes complicated with a manual definition of gates and regions, because different cell populations are present in the same fluorescence. CYTOGNOS recommends the use of the software INFINICYT for data analysis. Contact CYTOGNOS or its authorized distributor about the advantages and ways to get this analysis software.

To perform a manual analysis of the results we recommend follow the template shown in the following figures. Data show representative flow cytometry data on peripheral blood (healthy individual) stained with LYMPHOGRAM®.

- Dot plot n° 1: FSC versus SSC, ungated. Drawn a region (R1 or A) to select the lymphoid area according to its typical low light scatter properties. This region defines all lymphocytes and some contaminants.

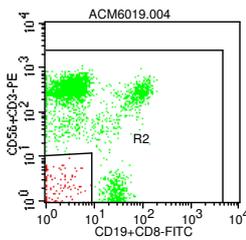
**1. Gate: No gate**



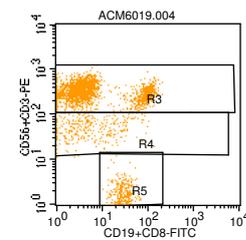
Gate Statistics			
File: ACM6019.004	Log Data Units: Linear Values		
Patient ID:	Tube: Untitled		
Acquisition Date: 18-Aug-05	Gate: No Gate		
Gated Events: 10000	Total Events: 10000		
X Parameter: FSC-Height (Linear)	Y Parameter: SSC-Height (Linear)		
Gate	Events	% Gated	% Total
G1	3103	31.03	31.03
G2	3209	32.09	32.09
G3	2055	20.55	20.55
G4	384	3.84	3.84
G5	748	7.48	7.48
G6	2988	29.88	29.88
T-LYMPHOCYTES	2006	20.06	20.06
NK CELLS	303	3.03	3.03
B-LYMPHOCYTES	679	6.79	6.79

- Dot plot n° 2: FL1 (CD19-FITC + CD8-FITC) versus FL2 (CD3-PE + CD56-PE), gated on region R1 or A. Drawn a region (R2 or B) to exclude all the events which does not express the markers included in FL1 and FL2.
- Dot plot n° 3: FL1 (CD19-FITC + CD8-FITC) versus FL2 (CD3-PE + CD56-PE), gated on G6 (total lymphocytes) defined as the events included in region R1 and R2 (R1 AND R2) or AB. Drawn a region (R3 or C) to include T lymphocytes subset (CD3+), a region (R4 or D) to include NK cells subset (CD3-CD56+), and a region (R5 or E) to include B lymphocytes subset (CD19+). With all this different region it can be defined the total T lymphocytes subset as the events included in the region R1 AND R2 AND R3 or ABC, the total NK cells subset as the events included in the region R1 AND R2 AND R4 or ABD, and the the B lymphocytes subset as the events included in the region R1 AND R2 AND R5 or ABE

**2. Gate: G1 = R1**



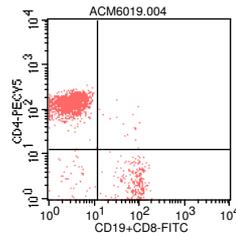
**3. Gate: G6 = R1 AND R2**



Region Statistics						
File: ACM6019.004	Log Data Units: Linear Values					
Patient ID:	Tube: Untitled					
Acquisition Date: 18-Aug-05	Gate: G6					
Gated Events: 2988	Total Events: 10000					
X Parameter: CD19+CD8-FITC (Log)	Y Parameter: CD56+CD3-PE (Log)					
Region	Events	% Gated	% Total	X Mean	Y Mean	
R1	2988	100.00	29.88	22.55	243.64	
R2	2988	100.00	29.88	22.55	243.64	
R3	2006	67.14	20.06	21.58	354.16	
R4	303	10.14	3.03	11.69	55.19	
R5	679	22.72	6.79	30.25	1.51	

- Dot plot n° 4: FL1 (CD19-FITC + CD8-FITC) versus FL3 (CD4-PECY5), gated on T lymphocyte subset defined as R1 AND R2 AND R3 or ABC. Set quadrants to distinguish between the T lymphocyte subsets: CD3+CD4+CD8- (upper left: UL), CD3+CD4+CD8+ (upper right: UR), CD3+CD4-CD8- (lower left: LL), CD3+CD4-CD8+ (lower right: LR).

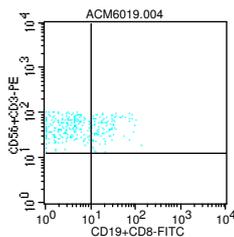
**4. Gate: T LYMPHOCYTES = R1 AND R2 AND R3**



Quadrant Statistics						
File: ACM6019.004	Log Data Units: Linear Values					
Patient ID:	Tube: Untitled					
Acquisition Date: 18-Aug-05	Gate: T-LYMPHOCYTES					
Gated Events: 2006	Total Events: 10000					
X Parameter: CD19+CD8-FITC (Log)	Y Parameter: CD4-PECY5 (Log)					
Quad Location: 12, 13						
Quad	Events	% Gated	% Total	X Mean	Y Mean	
UL	1505	75.02	15.05	3.45	148.93	
UR	16	0.80	0.16	58.04	62.45	
LL	55	2.74	0.55	3.57	2.79	
LR	430	21.44	4.30	85.98	2.16	

- Dot plot n° 5: FL1 (CD19-FITC + CD8-FITC) versus FL2 (CD3-PE + CD56-PE), gated on NK cell subset defined as the events included in region R1 AND R2 AND R4 or ABD. Set quadrants to distinguish between the NK cell subsets: CD3-CD56+CD8- (upper left: UL) and CD3-CD56+CD8+ (lower right: LR).

**5. Gate: NK CELLS = R1 AND R2 AND R4**



Quadrant Statistics						
File: ACM6019.004	Log Data Units: Linear Values					
Patient ID:	Tube: Untitled					
Acquisition Date: 18-Aug-05	Gate: NK CELLS					
Gated Events: 303	Total Events: 10000					
X Parameter: CD19+CD8-FITC (Log)	Y Parameter: CD56+CD3-PE (Log)					
Quad Location: 10, 12						
Quad	Events	% Gated	% Total	X Mean	Y Mean	
UL	202	66.67	2.02	3.41	54.78	
UR	101	33.33	1.01	28.27	56.03	
LL	0	0.00	0.00	***	***	
LR	0	0.00	0.00	***	***	

## RESULTS

The different lymphocyte subset counts can be expressed as the percentage of positive cells per lymphocytes or leucocytes present in the sample. Estimation of total lymphocyte percentage must be calculated as the events included in the regions R1 AND R2 or AB to avoid the contaminants included in the lymphoid area.

The different lymphocyte subsets count can be also expressed as the number of positive cells per microliter of sample (absolute counts). Absolute counts may be determined by two methods:

- The double platform method to calculate absolute counts combines results from hematology analyzer and flow cytometry and uses the following formula:  

$$\text{Absolute Counts (cells/}\mu\text{l)} = \text{Total White Blood Cell Count (cells/}\mu\text{l obtained from haematology analyzer)} \times \% \text{ lymphocytes} \times \% \text{ positively-stained cells} \div 10^4$$
- The single platform method has emerged as the method of the choice for absolute cell enumeration because comparative laboratory and external quality assessment studies have demonstrated that this methodology offers a lower intra- and inter-laboratory variation<sup>(9, 10)</sup>. Microbead-based technology consists of known amounts of fluorescent microbeads are admixed to a known volume of stained sample in a lyse-no-wash technique and the beads are counted along with cells. CYTOGNOS recommends the use of Perfect-Count Microspheres™ (CYT-PCM-50) to determine absolute counts using the following formula:

$$\text{Absolute Count (Cells/}\mu\text{l)} = \frac{\text{Total number of cells of interest counted}}{\text{Total number of Perfect-Count Microspheres™ counted}} \times \frac{\text{N}^{\circ} \text{ of Perfect-Count Microspheres /}\mu\text{l}}{\text{(value specified by manufacturer)}}$$

## LIMITATIONS

- Blood samples should be stored at 18-22°C and be tested within the 24 hours after they are obtained.
- It is advisable to acquire stained samples on the cytometer as soon as possible to optimize the results. Nonviable cells may stain nonspecifically. Prolonged exposure of whole blood samples to lytic reagents may cause white cell destruction and loss of cells from the target population.
- When using whole blood procedures, all red blood cells may not lyse under following conditions: nucleated red blood cells, abnormal protein concentration or hemoglobinopathies. This may cause falsely decreased results due to unlysed red blood cells being counted as leucocytes.
- Results obtained by flow cytometry may be erroneous if the cytometer laser is misaligned or the gates are improperly set
- Each laboratory should establish a normal range for lymphocyte subsets using its own test conditions. The data for the reagent's performance have been obtained from whole blood samples collected with EDTA as anticoagulant. The reagent's performance may be affected by the use of other anticoagulants.
- Certain patients may present special problems due to altered or very low number of certain cellular population.
- Cells separated from whole blood by means of density gradients may not have the same relative concentrations of cells as unseparated blood. This may be relatively insignificant for samples from individuals with normal white blood cell counts. In leucopenic patients, the selective loss of specific subsets may affect the accuracy of the determination.
- It is important to understand the normal pattern of expression of these antigens and its relation to the expression of other relevant antigens to carry out an adequate analysis<sup>(11-14)</sup>
- Abnormal states of health are not always represented by abnormal percentages of certain leucocyte populations. An individual who may be in an abnormal state of health may show the same leucocyte percentages as a healthy person. For this reason, it is advisable to use the test results in combination with other clinical and diagnosis data.

## EXPECTED VALUES

Each laboratory must establish its own normal reference ranges for lymphocyte subsets counting, since such values may be influenced by age, sex and race (15-17). The reference ranges for different lymphocyte subsets shown in the following table are expressed as the percentage of the lymphocyte subpopulations of the total lymphocyte count. These data were determined using the analysis software CYTORAMA- LYMPHOGRAM® at CYTOGNOS, and correspond to peripheral blood samples from normal adult between the ages of 18 and 60 years.

LYMPHOCYTE SUBSET	MEAN	MAX VALUE	MIN VALUE
%Total T lymphocytes (CD3+)	76.6	88.68	50.87
% T lymphocytes CD4+CD8-	48.1	67.11	24.43
% T lymphocytes CD4-CD8++	23.7	53.64	12.16
% T lymphocytes CD4-CD8-+	4.0	7.48	1.42
% T lymphocytes CD4+CD8+	0.9	6.02	0.14
%Total B lymphocytes (CD19+)	13.0	25.5	4.98
% NK cells (CD56+CD3-)	9.7	24.4	2.07

## QUALITY CONTROL

- To obtain optimum results it is advisable to verify the precision of pipettes and that the cytometer is correctly calibrated.
- The fluorochromes fluorescein isothiocyanate (FITC), R-phycoerythrin (PE) and R-phycoerythrin-cyanine 5 (PECy5) emit in different wavelengths but show a certain spectral overlapping which must be corrected by means of electronic compensation if combinations of different monoclonal antibodies are used conjugated with these fluorochromes. The optimum levels of compensation can be established by analysis in a dot-plot diagram of cells from healthy individuals stained with mutually exclusive monoclonal antibodies conjugated with the fluorochromes to be used in the test.
- To evaluate the non-specific binding of the reagent, an appropriated isotype control tube can be prepared.

## PERFORMANCE CHARACTERISTICS

### Reproducibility:

10 repeated measures from three peripheral blood samples representing high, medium and low lymphocyte counts were evaluated. In the following table are shown the mean percentage of the different lymphocyte subsets of the total lymphocytes count, the standard deviation and the coefficient of variation obtained for each of the three levels studied:

Lymphocytes level	% Total T lymphocytes (CD3+)	% Total T lymphocytes CD4+CD8-	% Total T lymphocytes CD4-CD8+	% B lymphocytes (CD19+)	% NK cells (CD56+CD3-)
High	83.39 ± 0.49 (CV=0.59)	43.19 ± 0.73 (CV=1.69)	30.32 ± 1.05 (CV=3.46)	11.53 ± 0.43 (CV=3.73)	5.08 ± 0.24 (CV=4.72)
Medium	81.10 ± 0.42 (CV=0.52)	48.00 ± 0.52 (CV=1.08)	30.37 ± 0.88 (CV=2.90)	14.42 ± 0.31 (CV=2.15)	4.48 ± 0.42 (CV=9.38)
Low	86.53 ± 0.59 (CV=0.68)	58.72 ± 0.88 (CV=1.50)	24.91 ± 0.59 (CV=2.37)	1.72 ± 0.17 (CV=9.88)	11.72 ± 0.54 (CV=4.61)

### Specificity:

The CD3 antigen is found on the cell surface of mature thymocytes and T lymphocytes in peripheral blood.

The CD4 antigen is found on a subpopulation of peripheral blood T lymphocytes, some thymocytes and some malignant cells of T-cell origin. Monocytes and macrophages show a weak expression of CD4. Normal B lymphocytes and granulocytes do not express surface CD4 antigen.

The CD8 antigen is found on a subpopulation of peripheral blood T lymphocytes, 60% of thymocytes and a limited number of malignancies of T-cell origin. Normal B lymphocytes, monocytes and granulocytes do not express surface CD8 antigen.

The CD19 antigen is found on the cell surface of normal and neoplastic B cells, being absent from T cells, monocytes and granulocytes.

The CD56 antigen is found on human peripheral blood NK cells and subsets of CD3+CD4+ and CD3+CD8+ T cells. Normal B lymphocytes, monocytes and granulocytes do not express surface CD56 antigen.

To assess cellular cross-reactivity, the CD3, CD4, CD8, CD19 and CD56 antibodies comprising the LYMPHOGRAM<sup>®</sup> were screened on normal human adult donor blood samples. Results consistently demonstrated that these antibodies react specifically with the appropriate lymphocyte populations.

### Accuracy

Main lymphocyte subset percentages obtained with LYMPHOGRAM<sup>®</sup> were compared with results obtained with the triple antibody combinations CD3-FITC/CD4-PE/CD8-PECy5 and CD45-FITC/CD56-PE/CD3-APC. Regression statistics reported in the following table indicate that the results are substantially equivalent in their reactivity on peripheral blood samples. Values are expressed in terms of percentage (%) of the total lymphocyte count.

	% Total T lymphocytes (CD3+)	% Total T lymphocytes CD4+CD8-	% Total T lymphocytes CD4-CD8+	% B lymphocytes (CD19+)	% NK cells (CD56+CD3-)
R <sup>2</sup>	0.98	0.99	0.99	0.99	0.90
Slope	0.95	10.95	1.10	1.00	0.82
Intercept	2.91	5.19	-0.96	0.61	1.05
Range	63.8 – 87.5	24.8 – 61.5	11.5 – 46.4	4.4 – 22.8	4.6 – 12.23
CCI	0.99	0.99	0.99	0.99	0.97

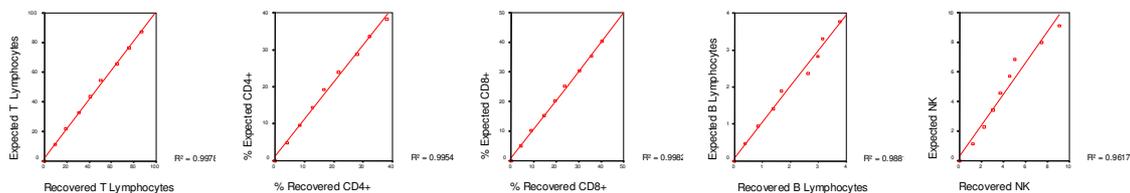
### Repetitivity

The results for 10 different samples stained with 2 different lots of LYMPHOGRAM<sup>®</sup> were compared. Each pair of data for the same sample was analyzed, obtaining the mean fluorescence intensity (MFI) and the standard deviation from which a grouped % CV was calculated. The results of the analysis are shown in the following chart:

	% Total T lymphocytes (CD3+)	% Total T lymphocytes CD4+CD8-	% Total T lymphocytes CD4-CD8+	% B lymphocytes (CD19+)	% NK cells (CD56+CD3-)
Mean	75.92	44.45	25.82	11.76	11.22
Grouped SD	2.35	2.07	1.19	1.23	0.99
Grouped CV (%)	3.10	4.66	4.61	10.46	8.82

### Linearity

Dilutions of a peripheral blood sample were made to check the concentration scale of stained cells obtained. The results show an excellent correlation level between the results obtained and expected based on the dilution used.



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#### EXPLANATION OF SYMBOLS

	Use by (use by YYYY-MM)
	Storage temperature limitation
	Consult instruction for use
	<i>In vitro</i> diagnostic medical device
	Batch code
	Code number
	Manufacturer

#### PRODUCED BY **CYTOGNOS SL**

Polígono La Serna, Nave 9  
37900 Santa Marta de Tormes  
Salamanca (España)  
Phone: + 34-923-125067  
Fax: + 34-923-125128  
Ordering information: [admin@cytoggnos.com](mailto:admin@cytoggnos.com)  
Technical information: [support@cytoggnos.com](mailto:support@cytoggnos.com)

[www.cytognos.com](http://www.cytognos.com)

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